

The Promise of Infrared Spectroscopy

Chandrajit Bajaj

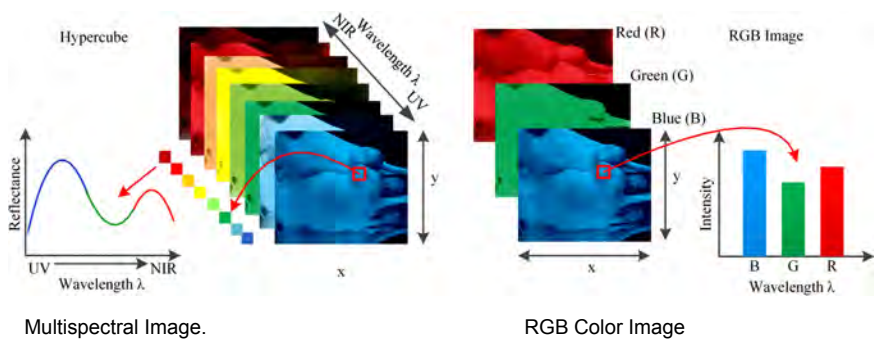
Sponsored in part by NIH- R01GM117594, NIH-R41GM116300 and a grant from SETON-Dell Medical 201602388



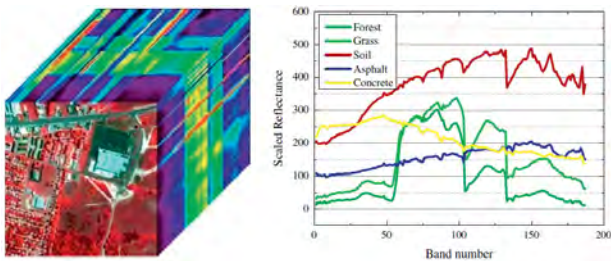
Computational Visualization Center (CVC) <http://cvcweb.ices.utexas.edu>
Dept. of Computer Science / Institute for Computational Engineering and Sciences
University of Texas at Austin

Diversity of Infrared Imaging

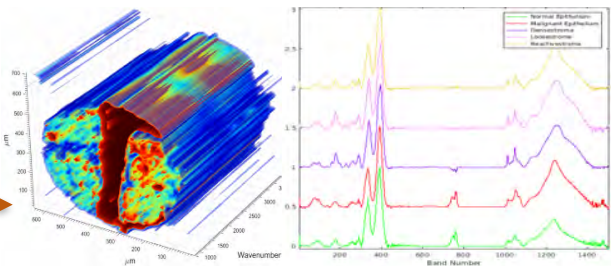
- Measures scalar intensities (absorbance, reflectance, transmittance, fluorescence,...) across various spectral channels



Region	Frequency f (THz)	Wavelength λ (μm)	Wavenumber ν (cm^{-1})
Visible	750 – 400	0.4 – 0.7	25000 – 14000
Near IR	400 – 200	0.7 – 1.4	14000 – 7000
SW IR	200 – 120	1.4 – 2.5	7000 – 4000
Mid IR	120 – 12	2.5 – 25	4000 – 400
Far IR	12 – 0.12	25 – 2500	400 – 4



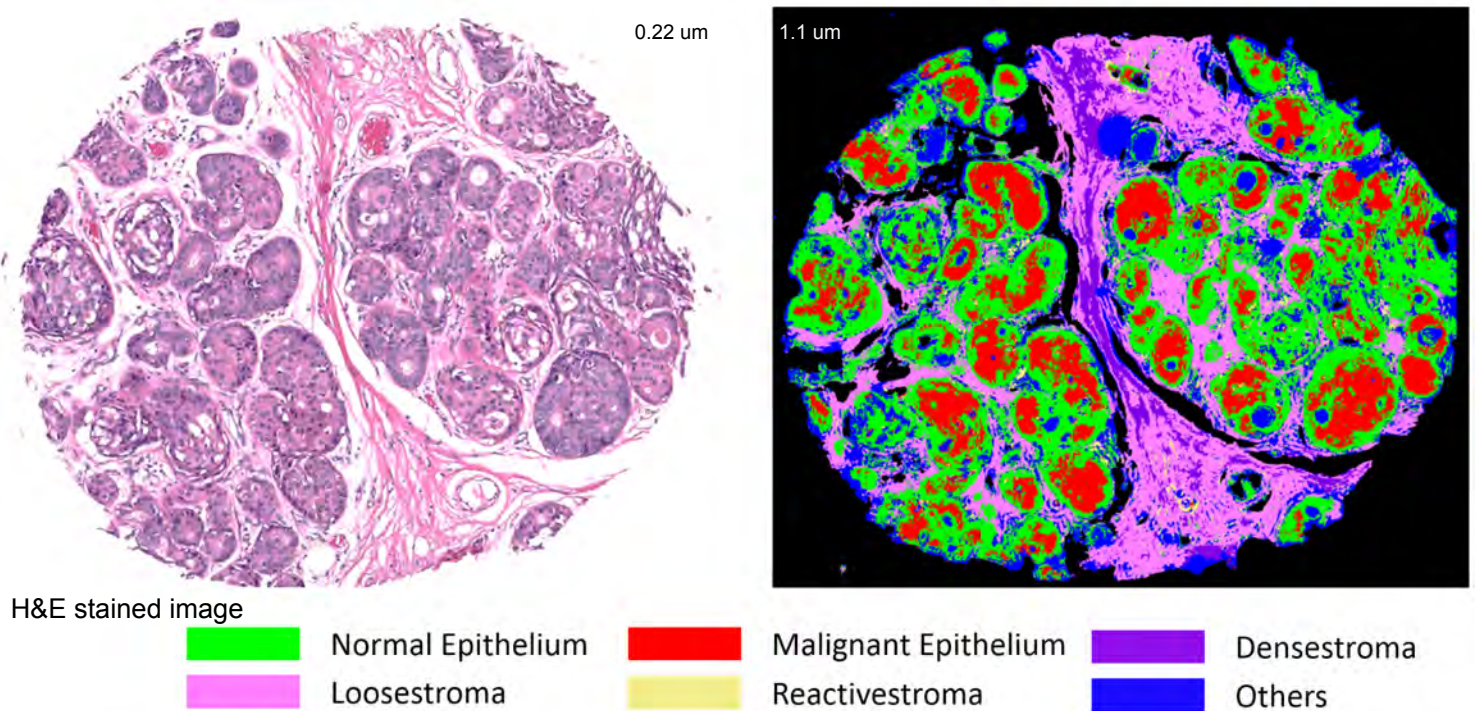
Hydrex HYMAP dataset: Resolution 5 m, Size 500 x 500, #Bands 128, Wavelength 0.4 -2.45 μm



Bottom: BR1003 AGILENT dataset: Resolution 1.1 μm ,² Size 128 x 128, #Bands 1506, Wavelength 2.5 -25 μm

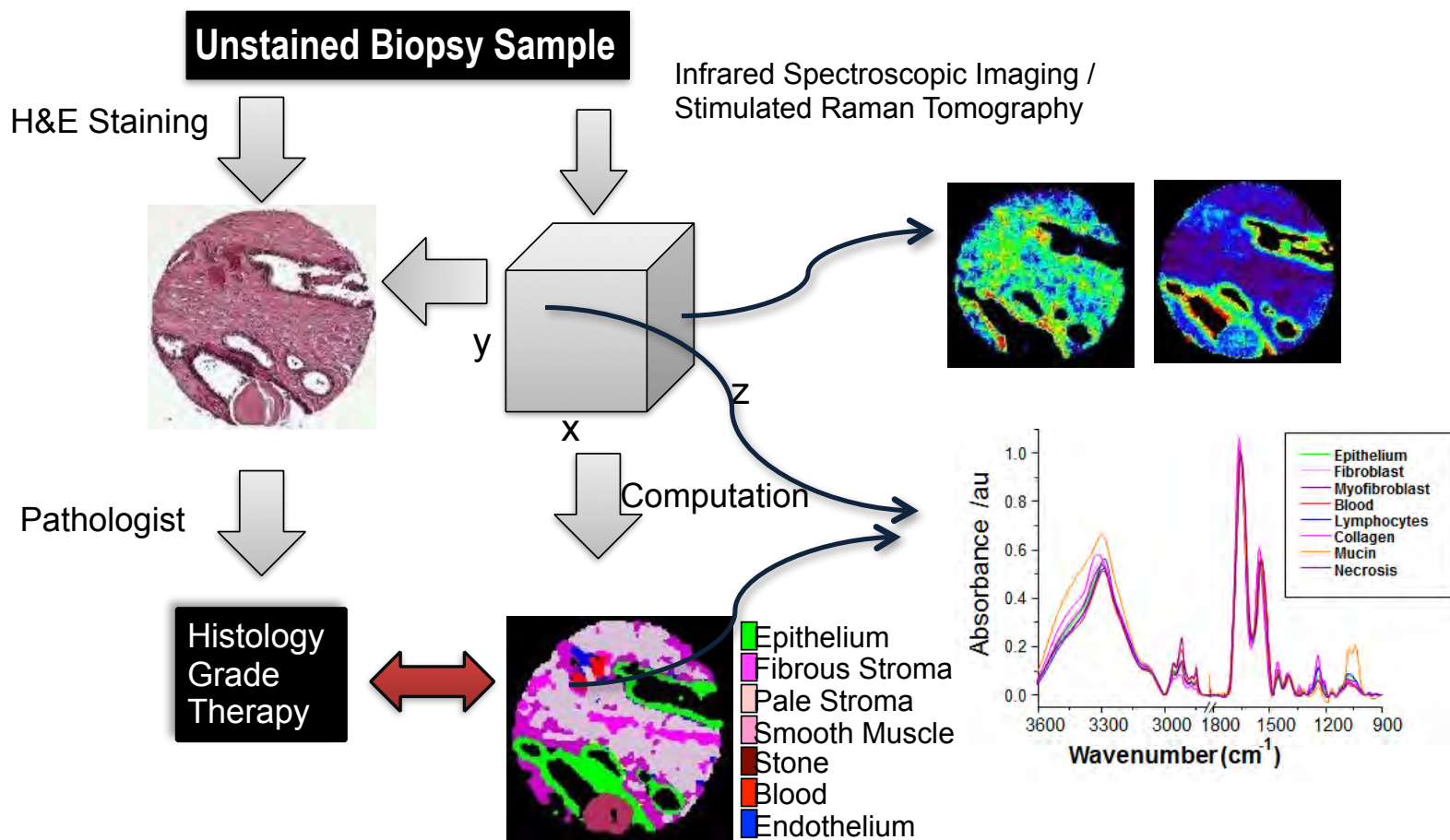
Visible vs Mid-Infrared

- Cell/Tissue Classification for Cancer Detection



Classified and pathologically verified map of the Tissue
Sample: Breast Tissue BR1003,

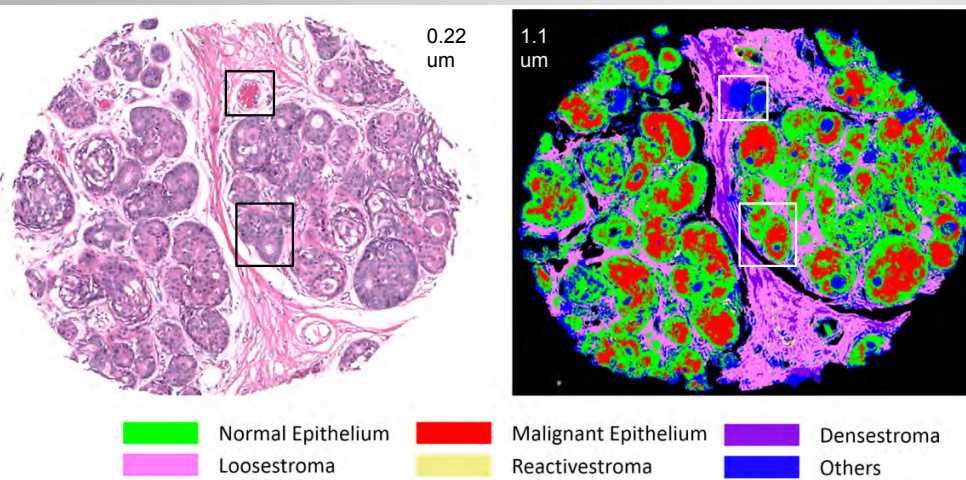
Tissue (Chemical) Imaging at Cellular Resolution



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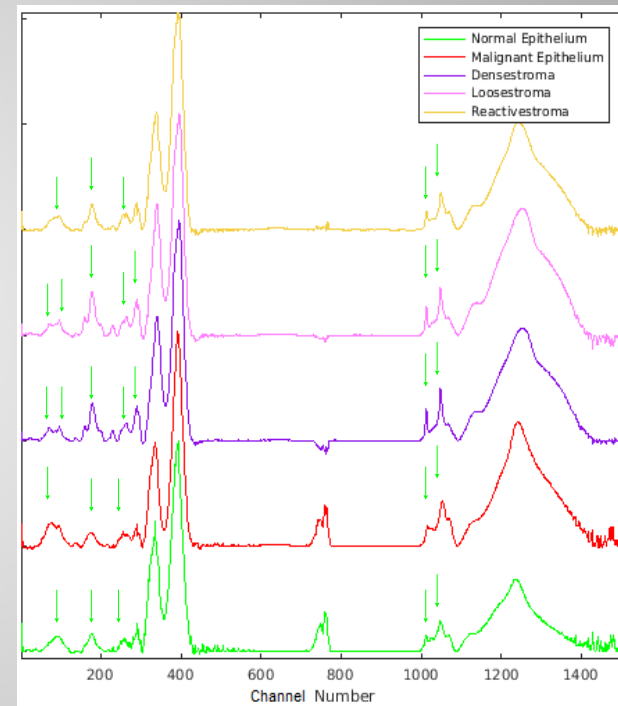
Small vs Big Data Spectral Analysis

Image Classification / Segmentation



0.005TB/day

1.3TB/day

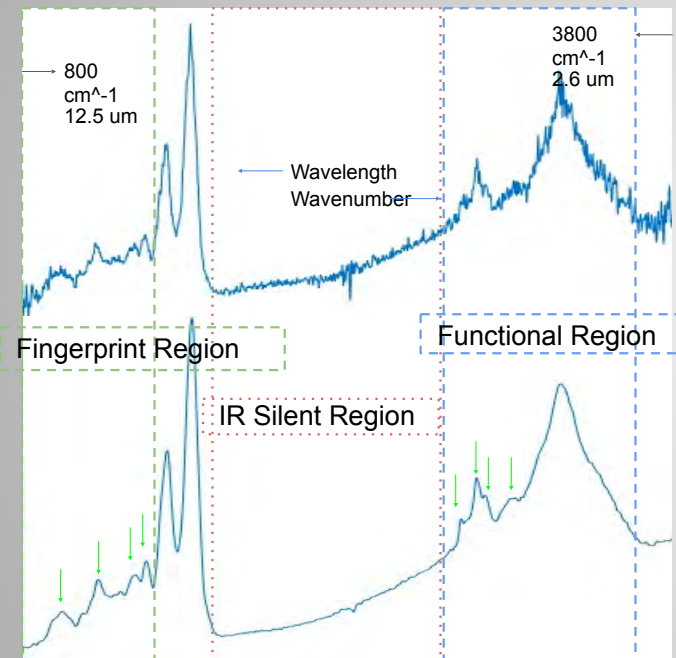


Typical spectra from five different classes of the core

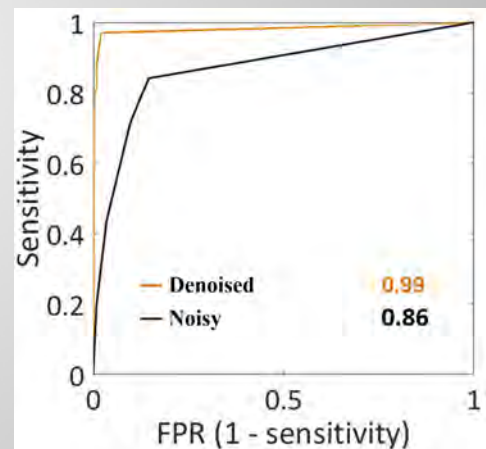
Need for Noise Estimation and De-Noising

Classification suffers if data is Noisy

- Classifying Malignant Epithelium using Machine Learning
- Denoised Signal: 98-99% correct prediction
- Noisy Signal: 85-86% correct prediction
- Based on well distinguished spectral markers

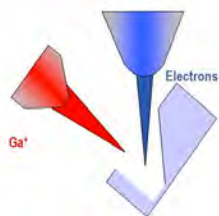
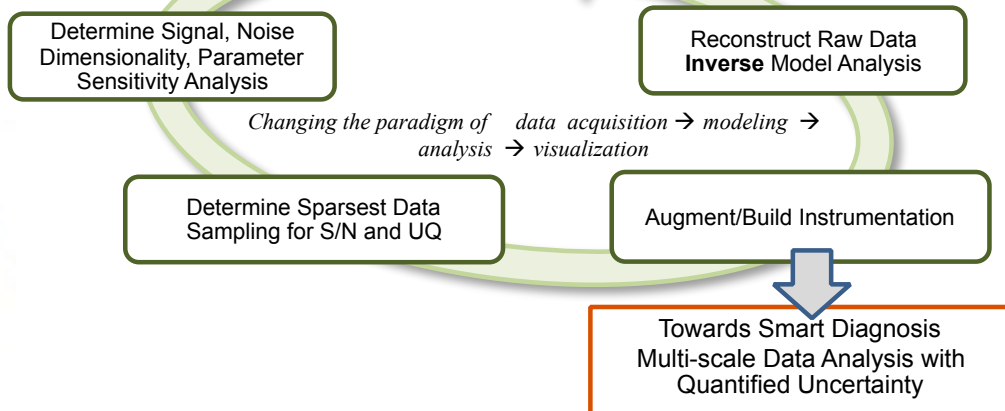
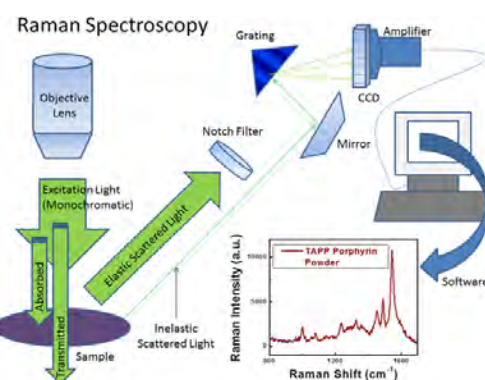
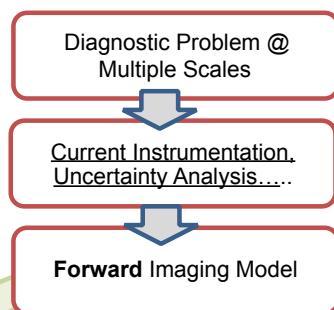
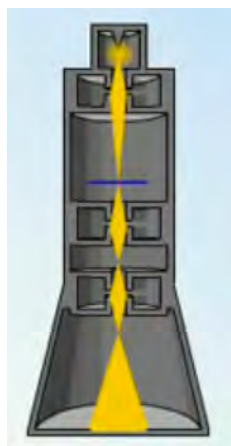


Noisy and DeNoised signal with spectral signature markers

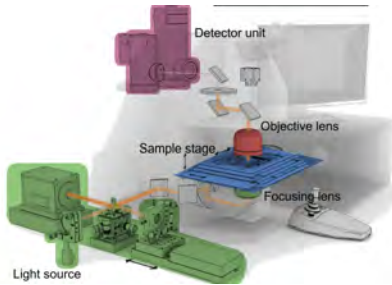


AUC curve for Malignant Epithelium
MNF: (Minimum Noise Fraction) a denoising technique

Combining IR Spectroscopy + Machine Learning



Noise Estimation: Model and Simulate the Image Acquisition



Goal: optimize for image quality

Given a collection Ψ of simulated samples with associated reference images $I_0(x, y, \tilde{\nu}; \mathcal{S})$ and probability distribution $\sum_{\mathcal{S} \in \Psi} p(\mathcal{S}) = 1$, determine the instrument configuration $\hat{\mathbf{c}}$ that gives the "best" expected image

$$\hat{\mathbf{c}} = \arg \min_{\mathbf{c}} \left(\sum_{\mathcal{S} \in \Psi} p(\mathcal{S}) \sum_x \sum_y [I(x, y, \tilde{\nu}; \mathcal{S}, \mathbf{c}) - I_0(x, y, \tilde{\nu}; \mathcal{S})]^2 \right)$$

- Minimize expected deviation from reference image I_0
- Reference samples $\mathcal{S} \in \Psi$ and associated images I_0 supplied by user
- Image function I computed using instrument model
- Using expected difference over collection of samples avoids overfitting
- Choice of sample collection allows optimization for certain types of samples

Computational flow of imaging forward model

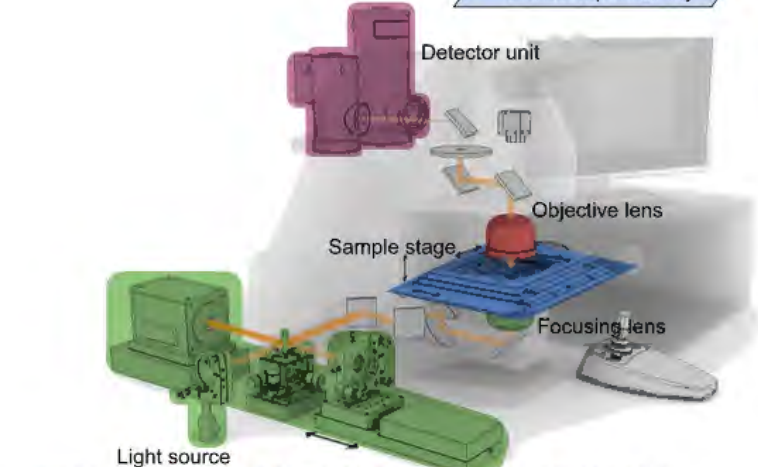
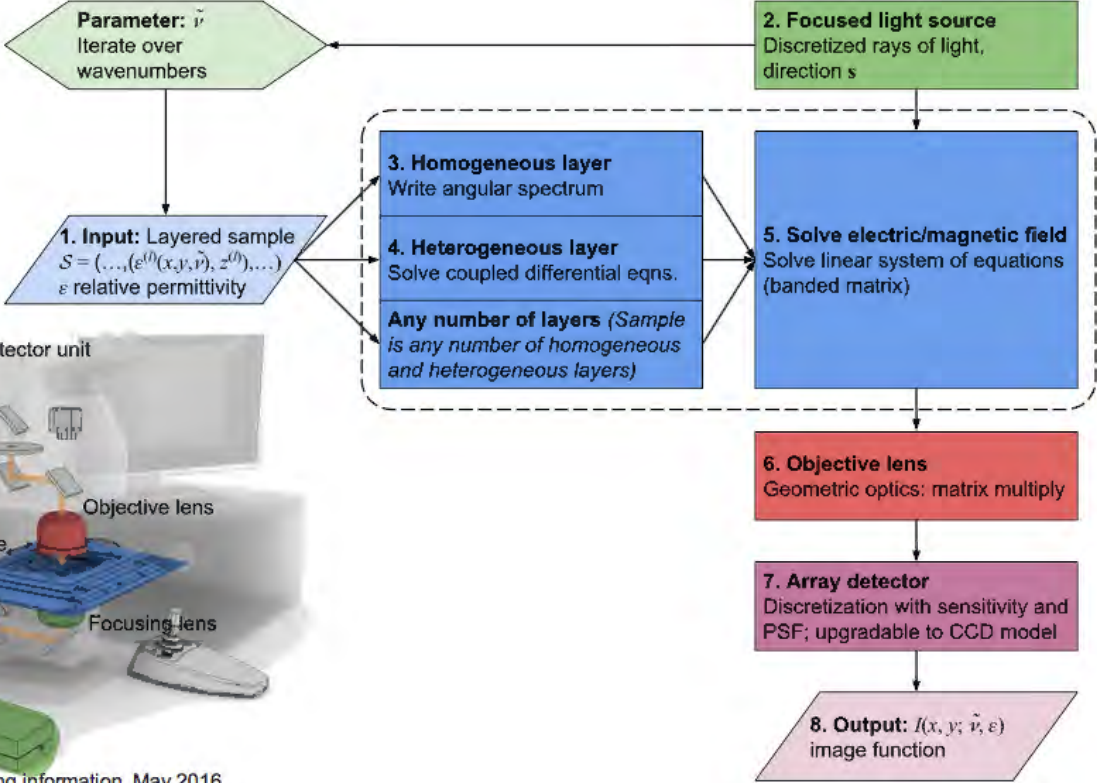
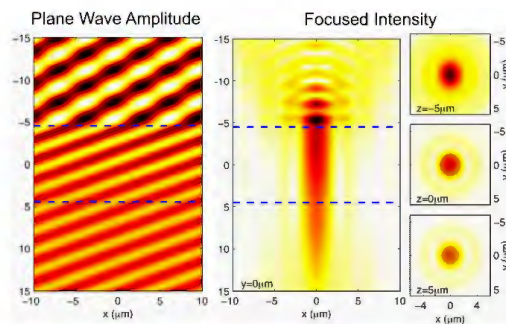
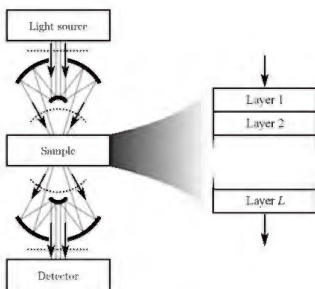
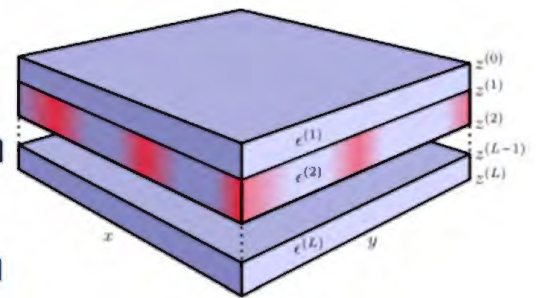
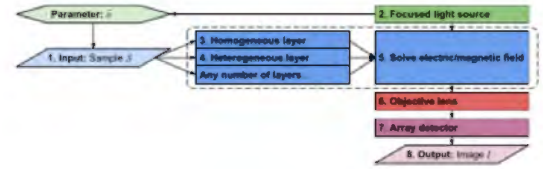


Image: Spegazzini. High definition (HD) IR imaging information. May 2016.



1. Input - Layered sample

- Model represents sample as layered medium
- Each layer may be modeled either homogeneously or heterogeneously
 - Homogeneous layers: $\epsilon^{(l)}(\tilde{\nu})$ depends only on wavenumber
 - Heterogeneous layers: $\epsilon^{(l)}(x, y, \tilde{\nu})$ depends on x, y position as well
- Variation along the z direction modeled through layers (discretization)
- Sample is described by relative permittivity ϵ — a four-dimensional $(l, x, y, \tilde{\nu})$ complex-valued function — and layer boundary positions $z^{(l)}$
- Sample $\mathcal{S} \approx ((\epsilon^{(1)}, z^{(1)}), (\epsilon^{(2)}, z^{(2)}), \dots, (\epsilon^{(L)}, z^{(L)}))$

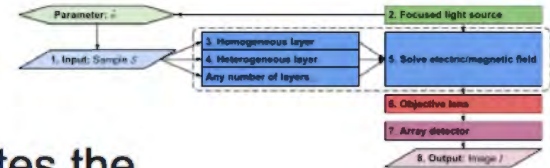
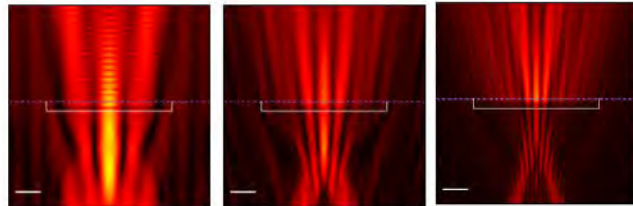
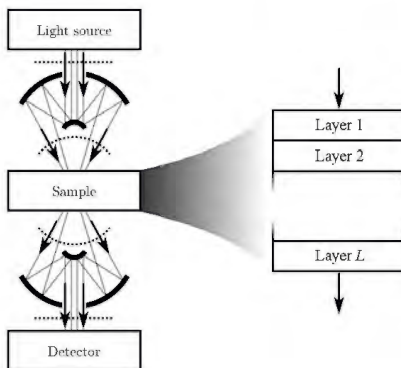


3-5. Light-sample interaction

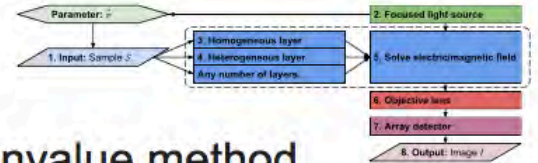
The sample interaction model (steps 3–5) computes the electric and magnetic field in the sample

- Generate constraint equations for each sample layer boundary
 - For homogeneous layers, get constraint equations directly (step 3)
 - For heterogeneous layers, solve differential system first (step 4)
- Solve linear system of $4(L - 1)N_F$ equations (step 5) to determine light leaving sample towards detector

$$E_{x,y}^{(l)}(x, y, z^{(l)}, \tilde{\nu}) = E_{x,y}^{(l+1)}(x, y, z^{(l)}, \tilde{\nu}) \quad H_{x,y}^{(l)}(x, y, z^{(l)}, \tilde{\nu}) = H_{x,y}^{(l+1)}(x, y, z^{(l)}, \tilde{\nu})$$



4. Heterogeneous layer computation



- System of differential equations solved via eigenvalue method

$$\begin{bmatrix} d\mathbf{X}(z, \tilde{\nu})/dz \\ d\mathbf{Y}(z, \tilde{\nu})/dz \\ d\mathbf{I}(z, \tilde{\nu})/dz \\ d\mathbf{J}(z, \tilde{\nu})/dz \end{bmatrix} = i2\pi\tilde{\nu}\Phi(\tilde{\nu}) \begin{bmatrix} \mathbf{X}(z, \tilde{\nu}) \\ \mathbf{Y}(z, \tilde{\nu}) \\ \mathbf{I}(z, \tilde{\nu}) \\ \mathbf{J}(z, \tilde{\nu}) \end{bmatrix}, \quad \Phi(\tilde{\nu}) = \begin{bmatrix} & & * & * \\ & & * & * \\ * & * & & \\ * & * & & \end{bmatrix} \in \mathbb{C}^{4N_F \times 4N_F}$$

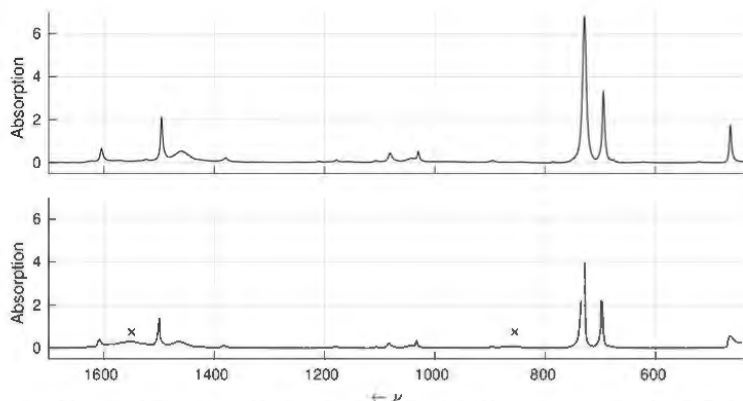
- Block-antidiagonal structure gives eigenvalues in opposite-sign pairs

$$0 = \det(\Phi - \lambda I) = \det \begin{bmatrix} -\lambda I & \Phi_1 \\ \Phi_2 & -\lambda I \end{bmatrix} = \det(\lambda^2 I - \Phi_1 \Phi_2)$$

- Solution is linear combination of eigenvalue pairs, eigenvectors
- Need eigenvalues, eigenvectors returned in opposite-sign pairs, not arbitrary order

Preliminary results: toluene

Top: Simulated by my implementation (solid line),
reference implementation (dotted line).
Bottom: Observed spectrum (Coblentz No. 10130).

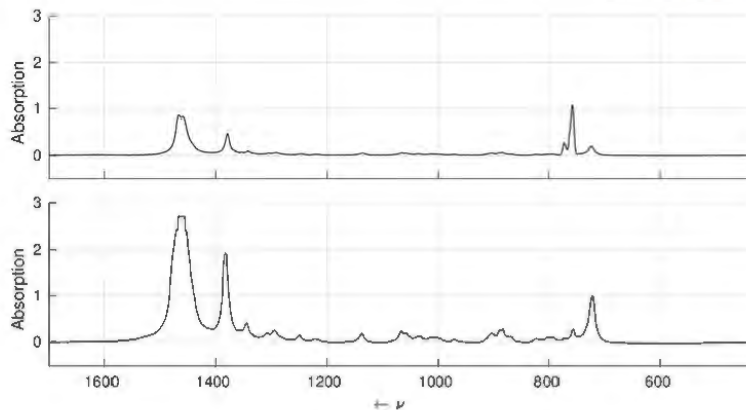


Coblentz Society, Inc. Evaluated infrared reference spectra. In Linstrom and Mallard, editors, *NIST Chemistry WebBook*, NIST Standard Reference Database Number 69. Gaithersburg MD, 20899. Accessed 30 May 2017.

SDBS No. 97. In *SDBSWeb: Spectral Database for Organic Compounds*. National Institute of Advanced Industrial Science and Technology, Tokyo. Accessed 5 June 2017.

Preliminary results: hexane

Top: output from my implementation (solid line),
reference implementation (dotted line).
Bottom: Observed spectrum (Coblentz No. 10118).

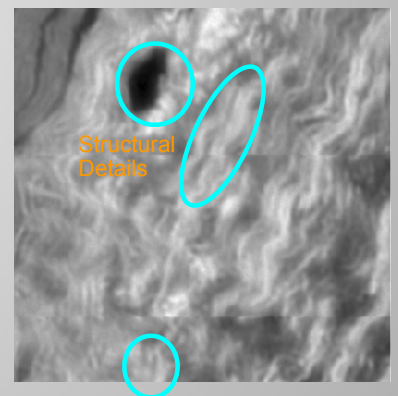
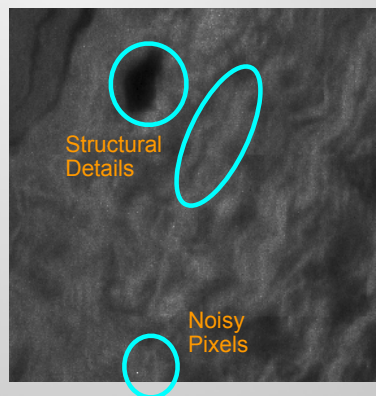
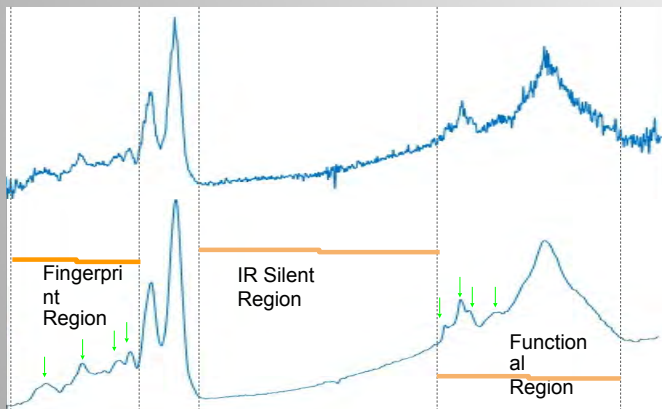


Coblentz Society, Inc. Evaluated infrared reference spectra. In Linstrom and Mallard, editors, *NIST Chemistry WebBook*, NIST Standard Reference Database Number 69. Gaithersburg MD, 20899. Accessed 30 May 2017.

The Spectral De-Noising Problem

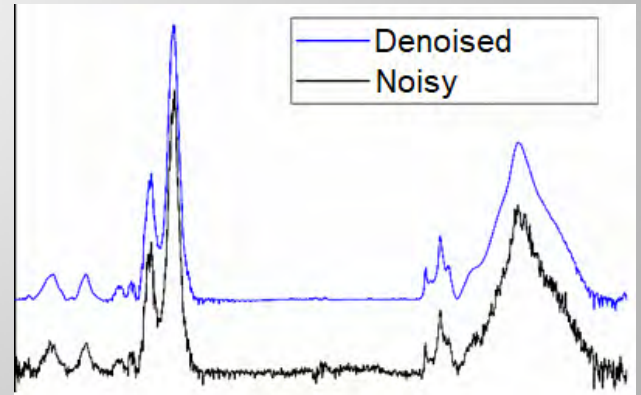
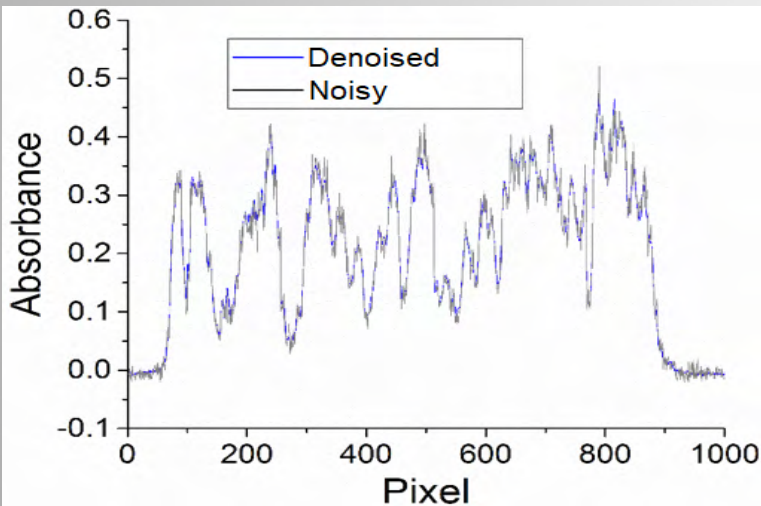
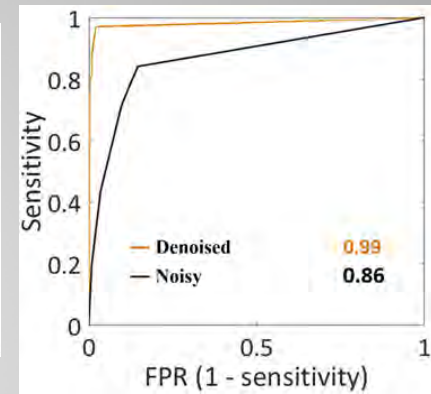
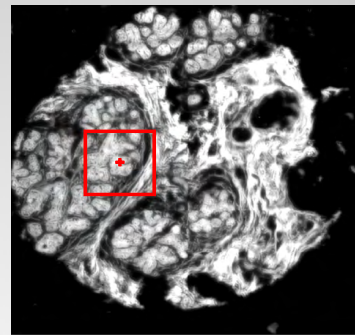
Given Noisy data ($Y = D + \epsilon$) under an unknown noise model ϵ , obtain an approximation \bar{D} :

1. Maximize the Signal to Noise Ratio (SNR) $\bar{D} = \underset{D}{\operatorname{argmax}} \frac{\operatorname{var}(D)}{\operatorname{var}(\epsilon)}$
2. Preserve structural details in the image domain
3. Preserve spectral markers F (peak position, peak location, relative peak spacing) $F = [j, L] \quad j \in [1 : S], L \in \{0, 1\}$



Metrics for De-Noising evaluation

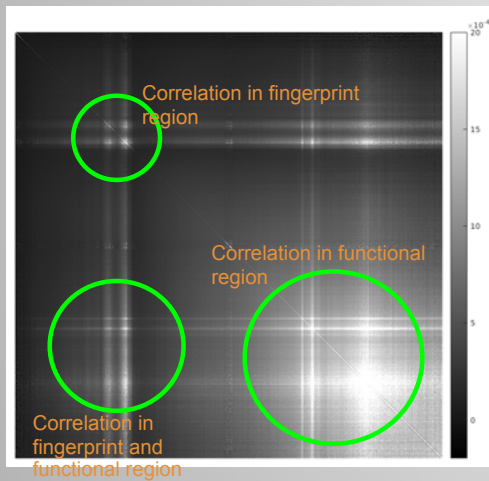
- Spatial
 - a. Classification Accuracy
- Spectral
 - a. Preservation of spectral features
 - b. Spatial profile analysis



Challenges for De-Noising in Mid IR

- Most algorithms fail if the data is noisy
- Spectral markers get corrupted or lost
- Signal and noise maybe correlated
- Noise correlated across multiple channels*

$$\epsilon_s \sim \rho \cdot \epsilon_{s-1} + \epsilon, \epsilon \sim \mathcal{N}(\mu, \sigma^2)$$



Covariance matrix of approximated noise in the FTIR data

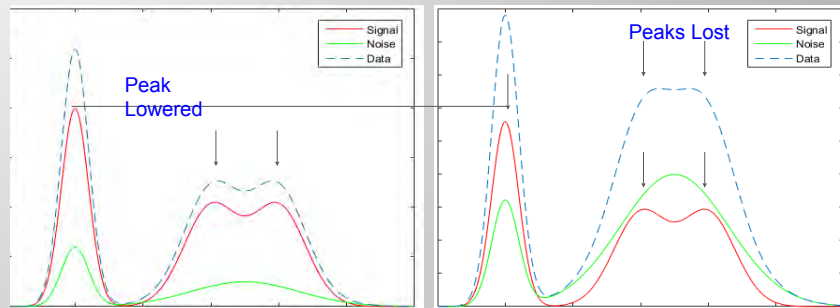
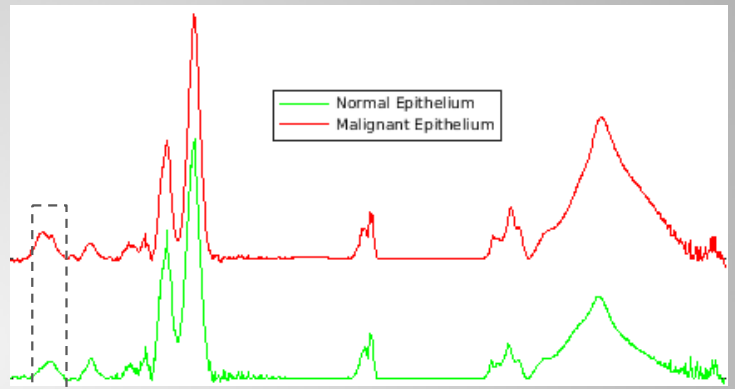
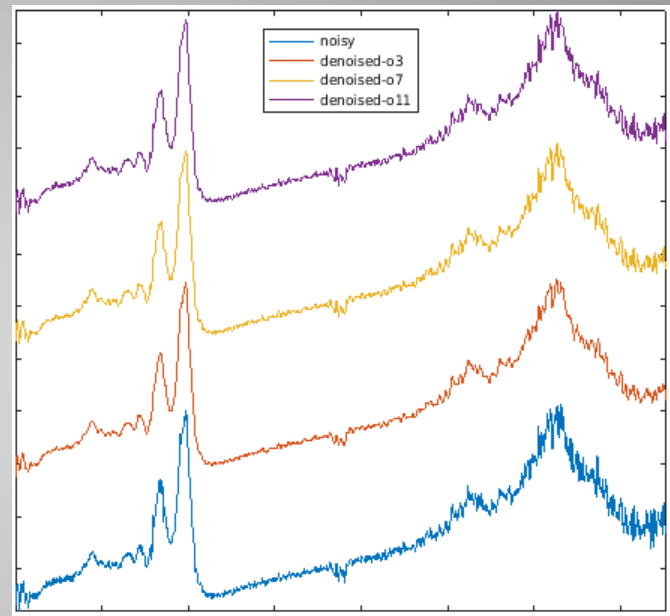
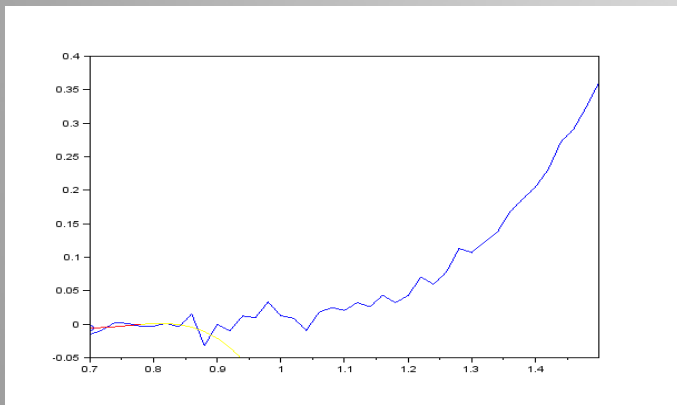


Illustration when noise is correlated within channels and also to data

* Leger et al. "Methods for systematic investigation of measurement error covariance matrices." *Chemometrics and Intelligent Laboratory Systems* 77.1 (2005): 181-205.

Prior De-Noising Techniques

- Filtering based approaches on spectrum *\$+
- Coefficients chosen by human experimentation
- Can handle any kind of noise
- Initial analysis is very time consuming
- Recalculate for every new dataset
- Good Classification accuracy



Results of Savitzky Golay filtering with different order polynomial fitting

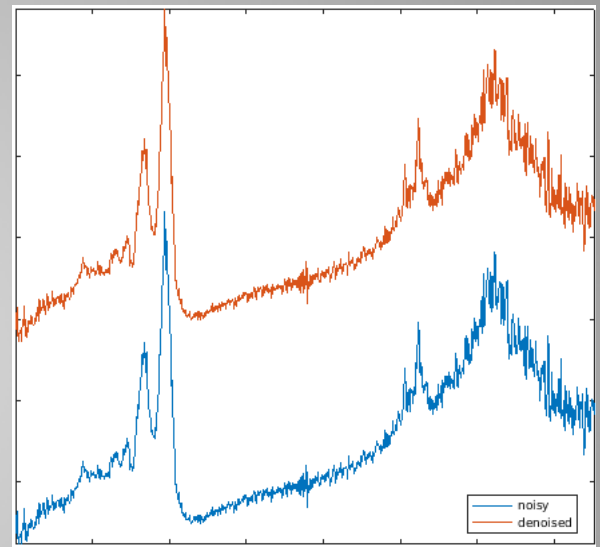
* Savitzky, Abraham, and Marcel JE Golay. "Smoothing and differentiation of data by simplified least squares procedures." Analytical chemistry 36.8 (1964): 1627-1639.

\$ Kawata, Satoshi, and Shigeo Minami. "Adaptive smoothing of spectroscopic data by a linear mean-square estimation." Applied spectroscopy 38.1 (1984): 49-58.

+ Tsai, Fuan, and William Philpot. "Derivative analysis of hyperspectral data." Remote Sensing of Environment 66.1 (1998): 41-51.

Prior De-Noising Techniques

- Signal is much stronger than noise
 - Signal and noise are independent
 - Noise across channels are independent
- Recover a low rank structure from the data where all the major signal contributions lie, and with sparse noise structure ^{*,+}
- Perform multi-modal iteration using Parallel Factor Analysis or Multidimensional Wiener based filters on the HSI cube ^{\$%}
- Use a rank-1 approximated tensor decomposition approach [#]
- Model the signal and noise as gaussian random fields and solve using Bayesian approach [&]



Assumption on independent noise model results in negligible denoising

$$\delta_i = \exp(-c \sum_{\delta} (i, i))$$

* Zhang et al. "Hyperspectral image restoration using low-rank matrix recovery." *IEEE Transactions on Geoscience and Remote Sensing* 52.8 (2014): 4729-4743

+ Wei, et al. "Hyperspectral image denoising via noise-adjusted iterative low-rank matrix approximation." *IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing* 8.6 (2015): 3050-3061

\$ Xuefeng et al. "Reduction of signal-dependent noise from hyperspectral images for target detection." *IEEE Transactions on Geoscience and Remote Sensing* 52.9 (2014): 5396-5411

% Letexier, Damien, and Salah Bourennane. "Noise removal from hyperspectral images by multidimensional Filtering." *IEEE Transactions on Geoscience and Remote Sensing* 46.7 (2008): 2061-2069

Guo, Xian, et al. "Hyperspectral image noise reduction based on rank-1 tensor decomposition." *ISPRS journal of photogrammetry and remote sensing* 83 (2013): 50-63.

& Zhong et al. "Jointly learning the hybrid CRF and MLR model for simultaneous denoising and classification of hyperspectral imagery." *IEEE Transactions on Neural Networks and Learning Systems* 25.7 (2014): 1319-1334

Minimum Noise Fraction (MNF)

- Developed by Green et al. *
- Can handle cases when signal and noise are uncorrelated
- Noise can be correlated within channels
- Orders data in terms of SNR in the MNF transformed space

Algorithm 1 MNF($Y \in \mathbb{R}^{N \times S}$)

- 1: Estimate Data Covariance Matrix: $\Sigma_Y = \text{Cov}\{Y\}$
 - 2: If δ is unknown, estimate: $\delta \sim Y(j, :) - Y(j+1, :), \forall j = 1 : N-1$
 - 3: Estimate Noise Covariance Matrix: $\Sigma_\delta = \frac{1}{2} \text{Cov}\{\delta\}$
 - 4: Calculate MNF projection vectors: $(\Sigma_Y \Sigma_\delta^{-1})\Phi = \Lambda_{MNF}\Phi$
 - 5: Project data along MNF vectors: $Y_{MNF} \in \mathbb{R}^{N \times S} = Y\Phi$
 - 6: Initialize $R \in \mathbb{R}^{S \times S}$ as top rank-K matrix
 - 7: Project data back: $\bar{D} \in \mathbb{R}^{N \times S} = Y_{MNF}R(\Phi^{-1}) = Y\Phi R(\Phi^{-1})$
-

* Green, Andrew A., et al. "A transformation for ordering multispectral data in terms of image quality with implications for noise removal." IEEE Transactions on geoscience and remote sensing 26.1 (1988): 65-74

Minimum Noise Fraction: Geometry

$$\begin{aligned}\Sigma_Y &= \Sigma_D + \Sigma_\delta && \text{Data Covariance Structure} \\ \Sigma_\delta &= E\Lambda E^T && \text{Spectral Decomposition of Noise Covariance} \\ \Sigma_W &= (E\Lambda_\delta^{-1/2})^T \Sigma_Y (E\Lambda_\delta^{-1/2}) \\ &= (E\Lambda_\delta^{-1/2})^T \Sigma_D (E\Lambda_\delta^{-1/2}) + (E\Lambda_\delta^{-1/2})^T \Sigma_\delta (E\Lambda_\delta^{-1/2}) \\ &= \Sigma_{W(D)} + I_\delta && \text{Whitened Noise Covariance} \\ \Sigma_W &= G\Lambda_{MNF}G^T && \text{Spectral Decomposition of Whitened Data}\end{aligned}$$

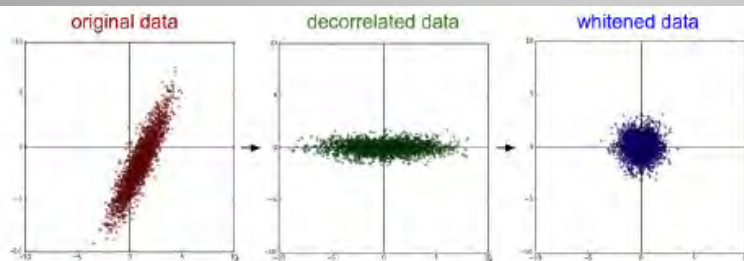


Figure: Geometric Interpretation of Whitening. Left: Orientation of noise. Middle: Orientation of noise after multiplication with E . Right: Orientation of noise after multiplication with $\Lambda_\delta^{-1/2}$

Minimum Noise Fraction: Fast MNF

$$D = Y * \Phi * R * \Phi^{-1}$$

$$= Y * (E * \Lambda_{\delta}^{-1/2} * G) * R * (E * \Lambda_{\delta}^{-1/2} * G)^{-1}$$

$$= Y * E * \Lambda_{\delta}^{-1/2} * G * R * G^T * \Lambda_{\delta}^{1/2} * E^T$$

$$= Y * E * \Lambda_{\delta}^{-1/2} * G * R * R^T * G^T * \Lambda_{\delta}^{1/2} * E^T$$

$$= Y * \hat{\Phi} * \tilde{\Phi}^T // \text{fastMNF}$$

$$\Rightarrow R = \left[\begin{array}{c|c} I_K & 0 \\ \hline 0 & 0 \end{array} \right]$$

$$\Rightarrow \hat{\Phi} = E * \Lambda_{\delta}^{-1/2} * G * R \quad // \text{forward MNF transform}$$

$$\Rightarrow \tilde{\Phi} = E * \Lambda_{\delta}^{1/2} * G * R \quad // \text{inverse MNF transform}$$

- Previously slow due to inversion of large matrices
- Current formulation avoids any inverse
- Uses rank-K approximation of G

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- 7: Project data back: $\tilde{D} \in \mathbb{R}^{N \times S} = Y_{MNF}R(\Phi^{-1}) = Y\Phi R(\Phi^{-1})$

Variations on Minimum Noise Fraction: Fast MNF

Approx MNF

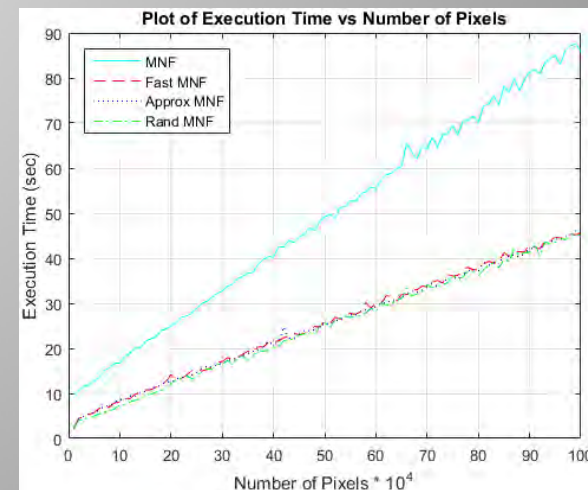
Rand MNF

- Block Lanczos # method to compute a rank K' SVD
- Guarantees $(1+\epsilon)$ Frobenius and $(1+\epsilon)$ spectral norm approximation
- Guarantees ϵ per vector norm approximation
- Although the Block Lanczos algorithm can attain machine precision
- Block Lanczos is rel.slow when the matrix is large
- A faster randomized and memory efficient version &
- Computes the K' -SVD up to $(1+\epsilon)$ Frobenius norm relative error
- Only keeps a $N \times O(K'/\epsilon)$ sketch in memory

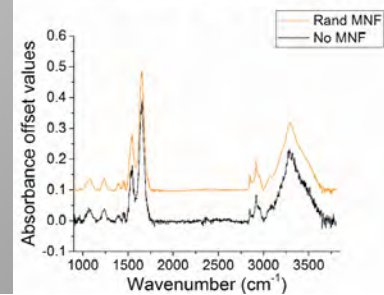
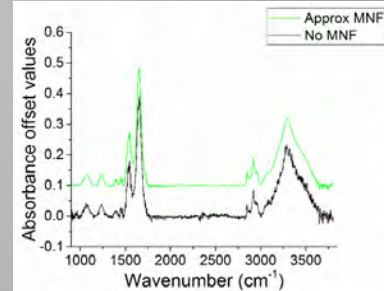
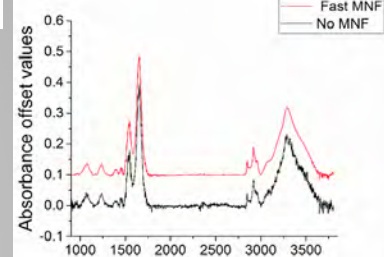
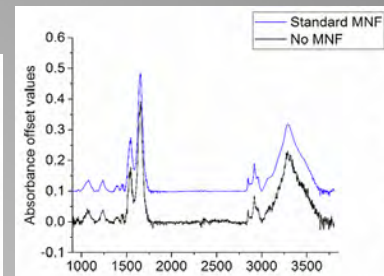
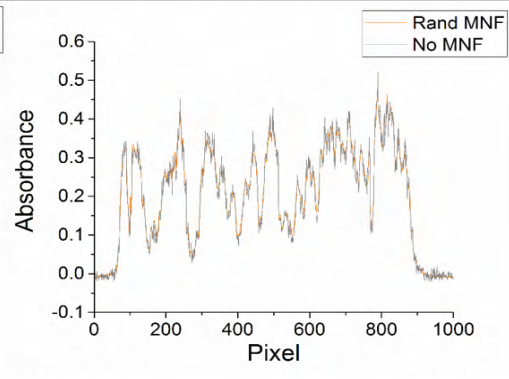
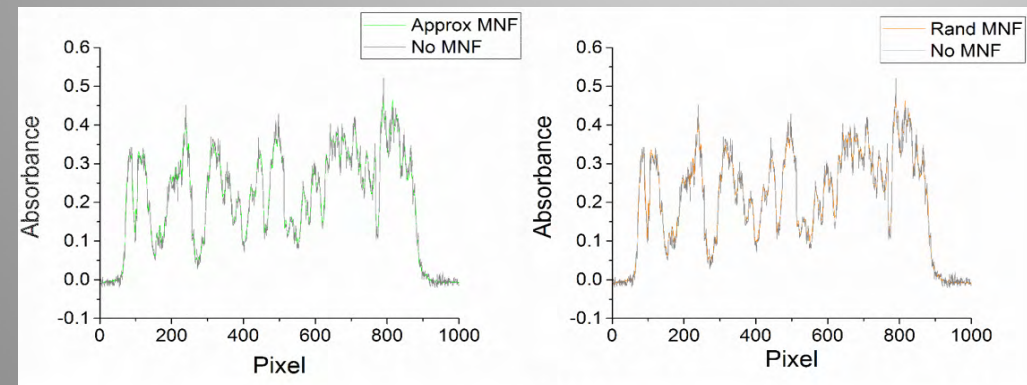
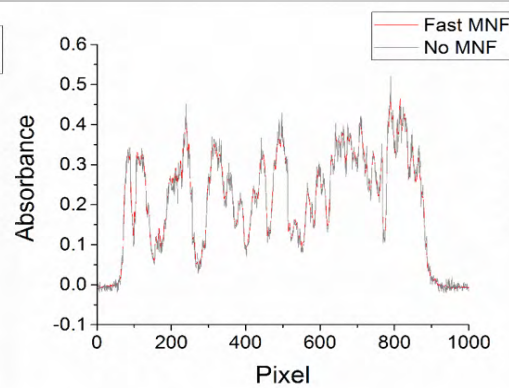
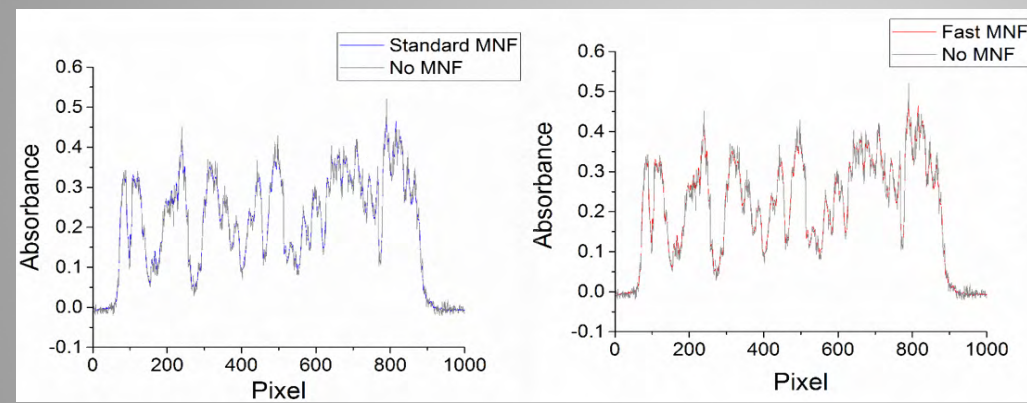
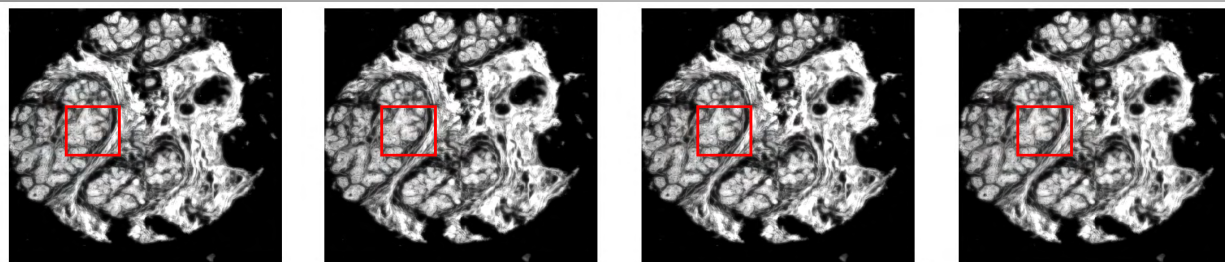
§ Musco, Cameron, and Christopher Musco. "Stronger approximate singular value decomposition via the block lanczos and power methods." Advances in Neural Information Processing Systems (NIPS) (2015).
 & Halko, Nathan, Per-Gunnar Martinsson, and Joel A. Tropp. "Finding structure with randomness: Probabilistic algorithms for constructing approximate matrix decompositions." SIAM review 53.2 (2011): 217-288.

Algorithm	Time	Space
1. MNF	$\mathcal{O}(S^3 + NS^2)$	$\mathcal{O}(NS + S^2)$
2. Fast MNF	$\mathcal{O}(S^3 + NSK)$	$\mathcal{O}(NK + S^2)$
3. Approx MNF	$\mathcal{O}(S^2K + NSK)$	$\mathcal{O}(NK + SK)$
4. Rand MNF	$\mathcal{O}(nnz(\Sigma_W)K + NSK)$	$\mathcal{O}(NK + SK)$

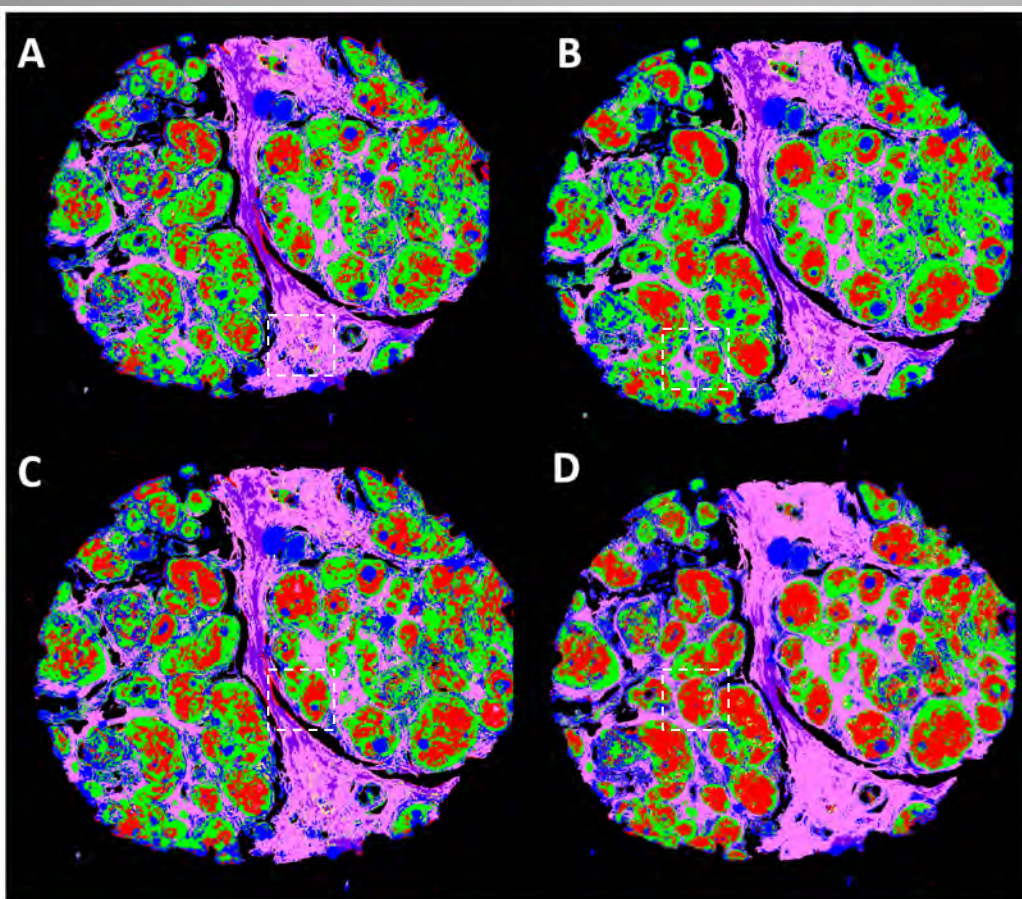
Table: Comparison of Time and Space complexity of MNF versions. \mathcal{O} : big-O complexity. $nnz(X)$: #non-zero elements in X . N : # pixels, S : # spectral bands and K : # chosen bands.



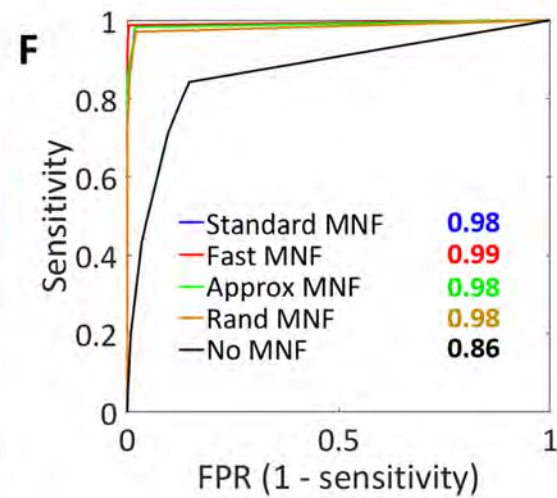
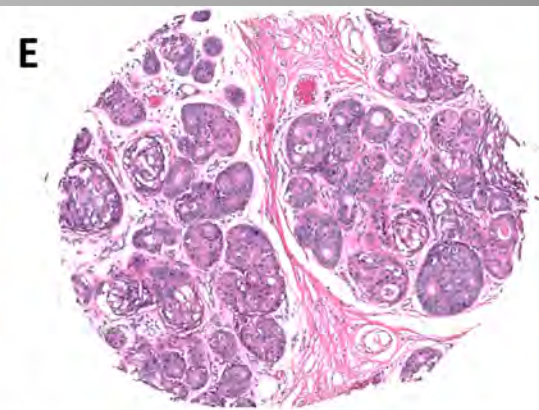
Minimum Noise Fraction: Profile Analysis



Minimum Noise Fraction: Classification Analysis



Normal Epithelium Malignant Epithelium Densestroma
Loosestroma Reactivestroma Others



Next Steps: Super-Resolution IR

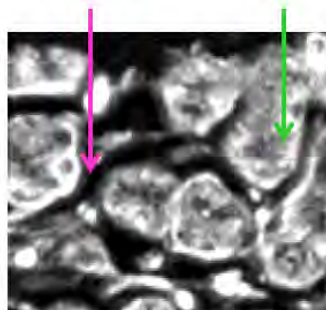
Breast Tissue

Visualization and chemical characterization of intralobular stroma.

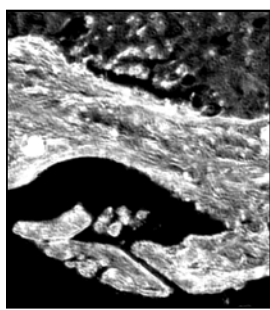
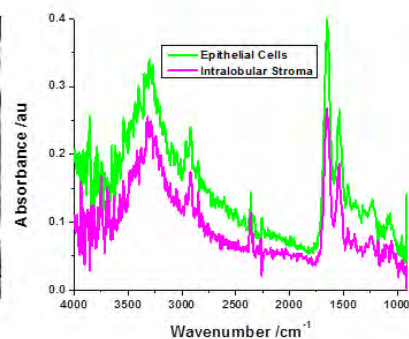


Global 15X
FT-IR

Intralobular Stroma Epithelial cells

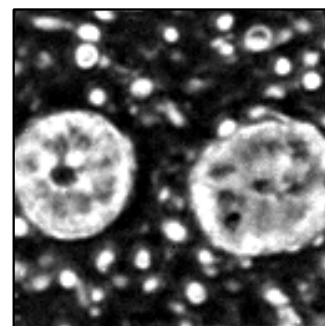
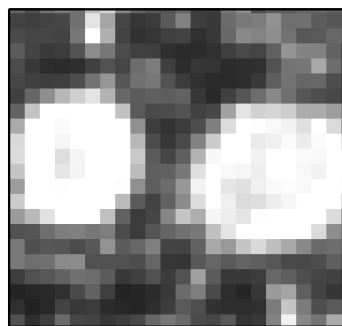


Global 74X
FT-IR



Prostate Tissue

Visualization and chemical characterization of collagen bands



Colon Tissue

Visualization and chemical characterization of subcellular mucin

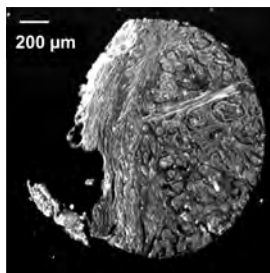
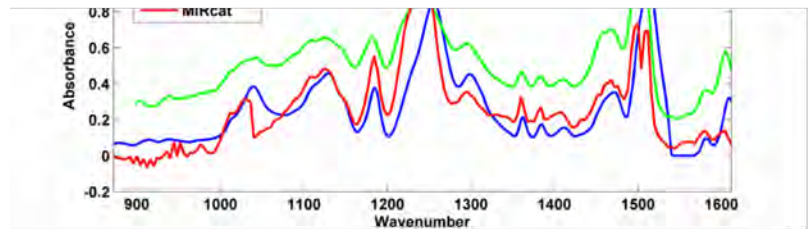
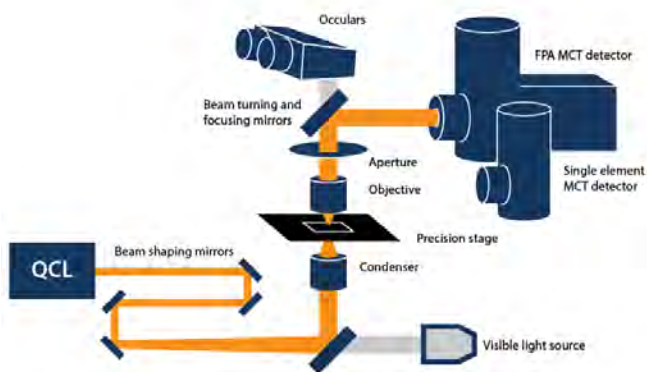
Computational Visualization Center (CVC) <http://cvcweb.ices.utexas.edu>
Dept. of Computer Science / Institute for Computational Engineering and Sciences
University of Texas at Austin

Nasse et al. *Nature Methods* (2011);
Reddy et al *Appl Spectrosc* (2013)

Collaboration with Rohit Bhargava, UIUC



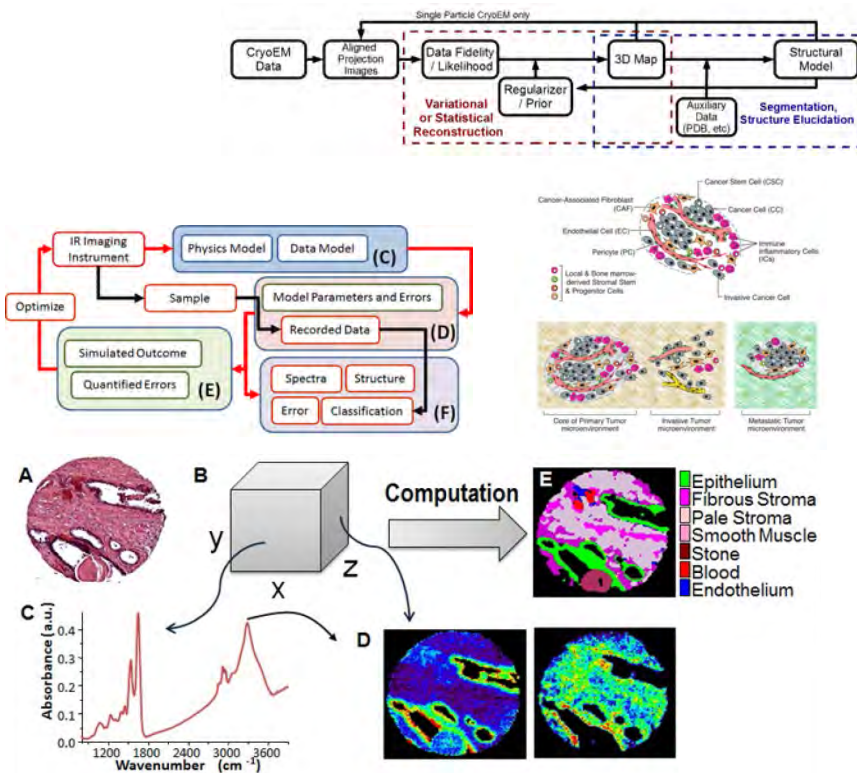
Next Steps: Tunable Quantum Cascade Laser (Compressed Sensing)



Frontier – Imaging with QCLs

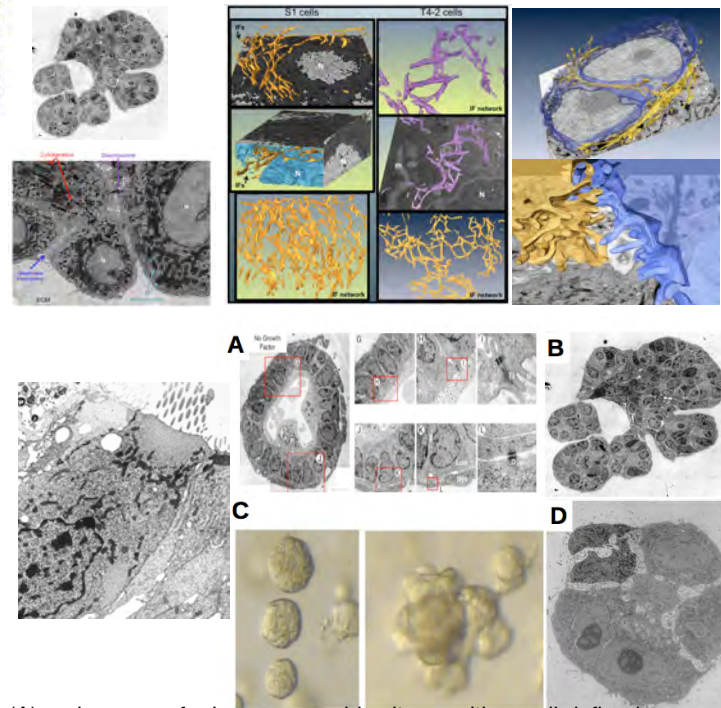
- Excellent SNR (10^4); 10^5 pix/s. $\Delta\nu$
- Ideal for ML approaches → use discrete frequency data for classification
- Limited frequency range

SMART DATA ANALYSIS: Tumor Cytotyping, Tracking Progression in 3D with Molecular-Cell Precision



(D) nucleic acids (left, at 1080cm⁻¹) and collagen specific (right, at 1245cm⁻¹).

Learning the Heterogeneity in cellular morphology, with a reduction in cell polarity during branching morphogenesis



- (A) acinar org. of primary organoid cultures with a well-defined lumen
- (B) 200 micron x 200 micron field of view of a single 100 nm thin resin section imaged at 1 nm resolution of an organoid undergoing branching morphogenesis
- (C) HMT-3522 S1 acini (left) and T4 aggregates (right).
- (D) S1 cells form growth-arrested acini with extensive intercellular membrane network and reduction in cell polarity, illustrating the label-free resolving power of the electron microscope.

Developing computational tools convert chemical imaging data to knowledge, as shown here for identifying all cells in prostate tissue (E).

DETAILS

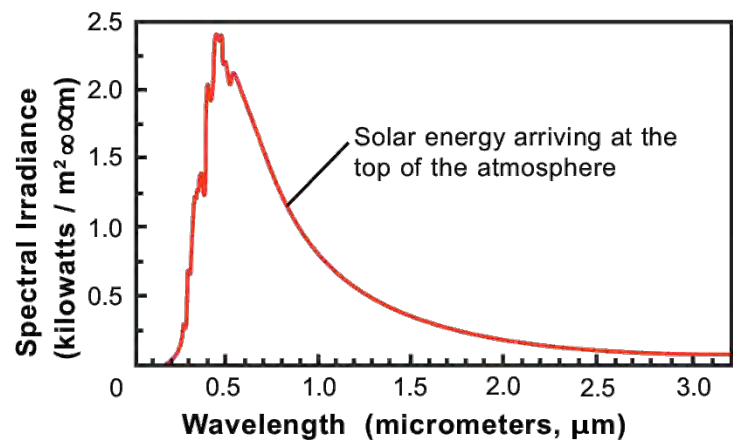
Sources for hyperspectral images

- Airborne Visible/Infrared Imaging Spectrometer (AVIRIS) program
 - Spatial resolution 20 meters at 20 km altitude, 4 meters at 4 km
 - 0.4-2.5 μm
- Advanced Spaceborne Thermal Emission and Reflection Radiometer (ASTER) Spectral Library
 - 2000 spectra of minerals, rocks, soils, water
 - 0.4-14 μm
- USGS Spectral Library
 - 500 spectra of minerals and a few plants
 - 0.2-3.0 μm

Reflectance vs. radiance

Laboratory instruments observe reflectance (or absorption), while remote sensing observes radiance, capturing several effects which must be corrected for.

- Spectrum of illuminating (solar) light
 - Additionally affected by shadows
- Light interactions with atmosphere
 - e.g. absorbance by water vapor, CO₂
- Illumination geometry (angle of incidence)
 - Varies by time of day and season
- Sensor characteristics
 - Variations between sensors, temporal changes



Hyperspectral image acquisition

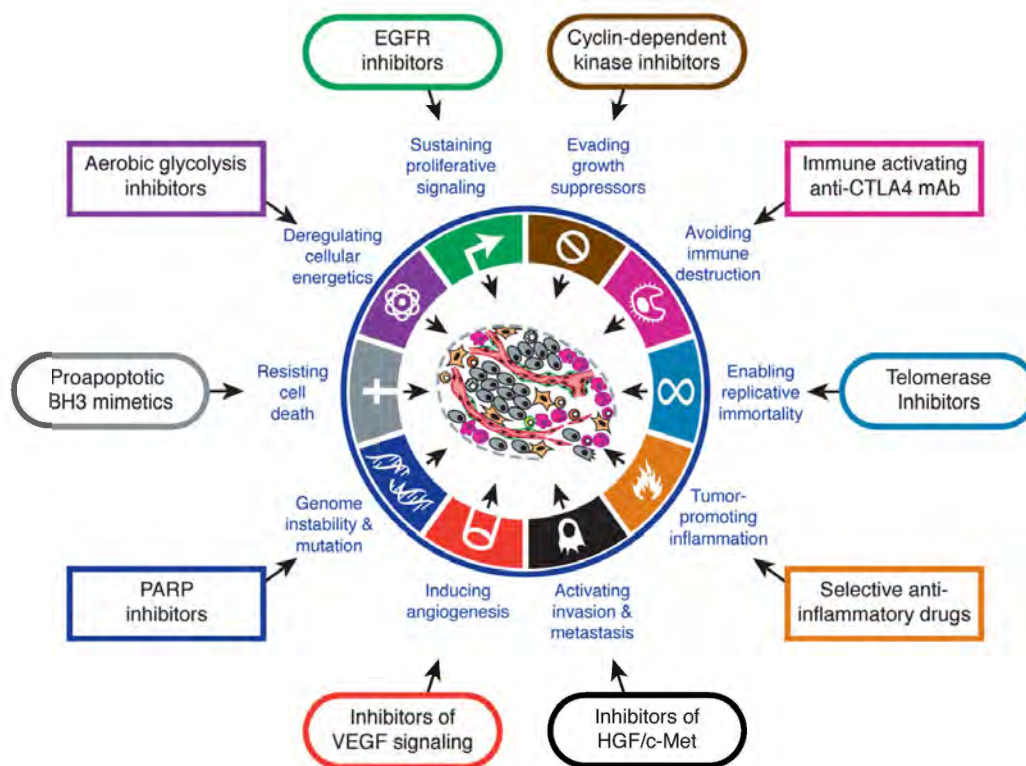
- Imaging spectrometers capture hyperspectral images
- Remote sensing
 - Analysis of the surface of the Earth (or other planets)
 - Optical dispersing element to separate frequencies
 - Can have spectral resolution as fine as $0.01\ \mu\text{m}$
 - Broadband (solar) light source
- Hyperspectral images also used in microscopy
 - Focal plane array detector
 - FTIR spectroscopy using interferometer light source

Hyperspectral instrument specifications

	AVIRIS	ASTER	Agilent Cary FTIR	QCL DFIR
Image size	677 pixels wide	4980 pixels wide (V/NIR)	128 × 128 pixels	128 × 128 pixels
Spatial resolution	20m (at 20km alt.) 4m (at 4km alt.)	15m (V/NIR) 30m (SWIR) 90m (MidIR)	1.1 μm	0.95 μm
Spectral resolution	0.01 μm in 0.4-2.5 μm	14 bands increments of 0.4-14 μm	0.0002 μm increments of 1.1-28.5 μm	0.002 μm in 5.25-12.87 μm
Detector	Si (Vis) InGaAs (NIR) InSb (SWIR)	Si (V/NIR) PtSi-Si (SWIR) HgCdTe (MidIR)	DLATGS or HgCdTe (MCT)	HgCdTe (MCT)
S/N	100 (at 0.40	varies	10,000	260 (single

Research and commercial imaging spectrometers

Sensor	Organization	Country	# bands	Wavelengths
AVIRIS	NASA	United States	224	0.4 - 2.5 μm
AISA	Spectral Imaging Ltd	Finland	286	0.45 - 0.9 μm
CASI	Itres Research	Canada	288	0.43 - 0.87 μm
DAIS 2115	GER Corp	United States	211	0.4 - 12.0 μm
HYMAP	Integrated Spectronics Pty Ltd	Australia	128	0.4 - 2.45 μm
DDORE	Earth Search Sciences	United	128	0.4 - 2.45 μm



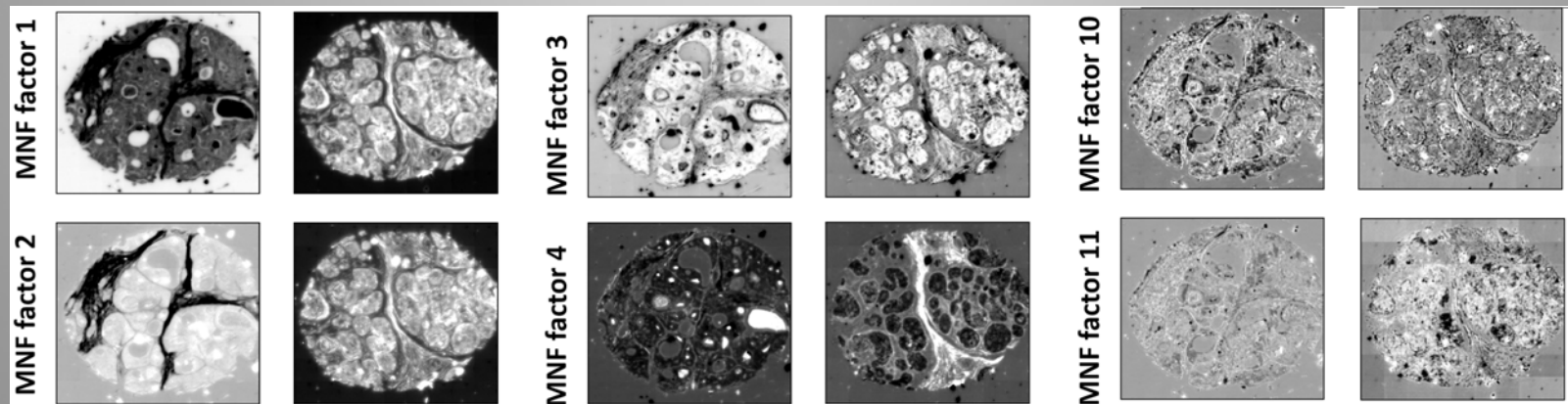
Minimum Noise Fraction: Automatic Band Selection

- Previous works include:
 - Manual inspection of Eigen-images
 - Automatic selection of Eigen-images based on its RMSE error to a clear image §
 - Computationally expensive
 - Time Consuming
- The optimal value of K can be determined from the diagonal entries of $\Lambda_{MNF} = SNR + 1$
- The Rose criteria & states that an SNR of at least 5.0 is needed to be able to distinguish image features at 100% certainty.

Algorithm 1 MNF($Y \in \mathbb{R}^{N \times S}$)

- 1: Estimate Data Covariance Matrix: $\Sigma_Y = \text{Cov}\{Y\}$
- 2: If δ is unknown, estimate: $\delta \sim Y(j, :) - Y(j+1, :), \forall j = 1 : N-1$
- 3: Estimate Noise Covariance Matrix: $\Sigma_\delta = \frac{1}{2} \text{Cov}\{\delta\}$
- 4: Calculate MNF projection vectors: $(\Sigma_Y \Sigma_\delta^{-1})\Phi = \Lambda_{MNF}\Phi$
- 5: Project data along MNF vectors: $Y_{MNF} \in \mathbb{R}^{N \times S} = Y\Phi$
- 6: Initialize $R \in \mathbb{R}^{S \times S}$ as top rank-K matrix
- 7: Project data back: $\hat{D} \in \mathbb{R}^{N \times S} = Y_{MNF}R(\Phi^{-1}) = Y\Phi(\Phi^{-1})$

§ Reddy, K., and Bhargava, R. "Accurate histopathology from low signal-to-noise ratio spectroscopic imaging data." Analyst 135.11 (2010): 2818-2825.
 & Bushberg et al. "The essential physics of medical imaging". Lippincott Williams & Wilkins, 2011.



Blind Error Metric

- Absence of ground truth
- The Method Noise Image (MNI)*
 - No-reference metric, simple and easy to use
 - Based on Structural Similarity Index Metric% (SSIM)
 - Scores based on intensity, contrast, Image moments
 - Maximum score around highly structured regions

Algorithm 4 MNI Metric (I, \hat{I})

```

1: Input:
2: Noisy Image  $I$ , Denoised Image  $\hat{I}$ 
3: Body:
4: Compute difference image  $\hat{I}$ :  $M = I - \hat{I}$ 
5: Compute  $SSM_P = SSIM(I, M)$ 
6: Compute  $SSM_Q = SSIM(\hat{I}, I)$ 
7: Compute image quality score  $e = Corr(SSM_Q, SSM_P)$ 
    
```

* Kong, Xiangfei, et al. "A new image quality metric for image auto-denoising." Proceedings of the IEEE International Conference on Computer Vision. 2013.

% Wang, Zhou, et al. "Image quality assessment: from error visibility to structural similarity." IEEE transactions on image processing 13.4 (2004): 600-612.

